## pAM\_dCas9\_CTCF Vector



Catalog No.: 53124
Format: 10 μg

## Description

The pAM\_dCas9\_CTCF Vector is designed for use with Active Motif's enChIP Kit (Catalog No. 53125). The enChIP method relies on the CRISPR/Cas9 system to direct a guide RNA (gRNA) to a specific genomic locus for immunoprecipitation. The gRNA is designed as the complementary sequence of the desired target locus. An enzymatically inactive form of the *Streptococcus pyogenes* Cas9 protein, which contains Active Motif's unique AM-tag sequence, is co-expressed with the gRNA. The gRNA directs the dCas9 protein to its target sequence, immediately upstream of a Protospacer Adjacent Motif (PAM) (5 ´- NGG). Recognition of a PAM site leads to unwinding of the DNA and formation of an RNA-DNA heteroduplex. Cells are then formaldehyde fixed and chromatin is prepared. An antibody directed against the AM-tag is used to enrich for genomic sequences bound by the gRNA/dCas9 complex. DNA can be analyzed by qPCR or NGS to identify the enriched genomic regions. The pAM\_dCas9\_CTCF Vector can be used as a positive control for the enChIP Kit. It contains a gRNA sequence corresponding to a CTCF binding site on chromosome 19 (5,804,115-5,804,209) which has been cloned into the pAM\_dCas9 empty vector (Catalog No. 53122). The pAM\_dCas9\_CTCF vector should be used with the single vector transfection protocol.

#### **Contents**

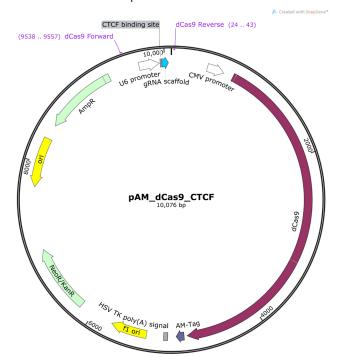
• 10 μg of pAM\_dCas9\_CTCF Vector provided at a concentration of 100 ng/μl.

## pAM dCas9 CTCF Vector Features and Circle Map

The following features are present in the pAM dCas9 CTCF Vector based on nucleotide sequence.

| dCas9 reverse primer  | 24-43      |
|---|------------|
| CMV promoter  | 430-633    |
| dCas9 coding region   | 748-4851   |
| AM-Tag  | 4888-4980  |
| Herpes simplex virus TK poly (A) signal                     | 5079-5127  |
| f1 origin of replication                                    | 5329-5757  |
| Neomycin <sup>R</sup> /Kanamycin <sup>R</sup> coding region | 6167-6961  |
| SV40 polyadenylation signal                                 | 7137-7258  |
| ColE1-derived plasmid origin of replication                 | 7709-8297  |
| β-lactamase (Amp <sup>r</sup> ) coding region               | 8468-9328  |
| dCas9 forward primer  | 9538-9557  |
| U6 promoter   | 9699-9939  |
| CTCF binding site   | 9949-9967  |
| gRNA scaffold   | 9969-10044 |
|   |            |

**Note:** The pAM\_dCas9 empty vector (Catalog No. 53122) is available separately and is recommended for use in combination with pAM\_dCas9\_CTCF as a no gRNA negative control.



#### **Quality Control**

Plasmid construct has been confirmed by restriction analysis and sequence verified. For the complete pAM\_dCas9\_CTCF Vector sequence, please visit the Documents tab at www.activemotif.com/enchip.

#### **Shipping & Storage**

Products are shipped on dry ice.

Resuspended DNA is stable for 6 months when stored at -20°C. Avoid repeated freeze/thaw cycles.



# Transfection of pAM\_dCas9\_CTCF expression plasmid

Cells can be transiently transfected with the pAM\_dCas9\_CTCF expression plasmid. The following protocol provides recommendations for transfection using FuGENE® HD Transfection Reagent (Catalog No. 32042). Optimization may be required for each cell line tested.

To determine the efficiency of the transfection, set up a duplicate transfection for each cell type used. Cell lysates can be prepared from the duplicate transfection reaction using Active Motif's Nuclear Extract Kit (Catalog No. 40010) for analysis by Western blot. Use the AM-Tag polyclonal antibody (Catalog No. 61677) at a 1:250 - 1:1,000 dilution for detection of the AM-tagged dCas9 protein. If the tagged protein is not detected, continue to optimize transfection conditions.

Calculate the number of transfections you will perform. Small scale reactions provide enough chromatin to perform one enChIP reaction. Large scale reactions provide enough chromatin to perform 5 enChIP reactions. We recommend running a no gRNA negative control reaction (empty pAM\_dCas9 vector, Catalog No. 53122) for each cell line tested.

1. Seed cells in either a 6-well plate (small scale) or 10cm dish (large scale) using the appropriate growth medium. Incubate in a humidified incubator for 24 hours. Cells should be 70-80% confluent at the time of transfection.

| Small Scale Cell Culture<br>(1 enChIP rxn) |                           |                           |  |
|--|---------------------------|---------------------------|--|
| Single Vector                              |                           | Single Vector             |  |
| Cell culture plate                         | 6-well plate              | 10 cm dish                |  |
| Cell seeding density•                      | 5 x 10 <sup>s</sup> cells | 3 x 10 <sup>6</sup> cells |  |
| Growth medium                              | 10 ml                     | 20 ml                     |  |

<sup>\*</sup>These conditions were established for cells with doubling times of 14-18 hours. Cell seeding densities may need to be optimized to ensure that cells are -80% confluent at the time of transfection.

2. Prepare a separate microcentrifuge tube for each transfection reaction. To each tube add the recommended amount of DNA and Opti-MEM according to the table below.

|                               |                     | Small Scale Cell Culture<br>Single Vector | Large Scale Cell Culture<br>Single Vector |
|-------------------------------|---------------------|---|---|
| Positive control<br>CTCF gRNA | (dCas9_CTCF) vector | 1 µg                                      | 2 μg                                      |
|                               | Opti-MEM            | Up to 275 μl                              | Up to 550 μl                              |
|                               |                     |   |   |
| No gRNA<br>negative control   | dCas9 vector        | 1 µg                                      | 2 μg                                      |
|                               | Opti-MEM            | Up to 275 μl                              | Up to 550 μl                              |

3. Add FuGENE HD Transfection reagent drop wise directly to the DNA/media mixture. Do not allow FuGENE to come directly in contact with the plastic from the tube. Mix the solution by pipetting up and down and incubate at room temperature for 30 minutes.

Small scale cell culture: Add 6 µl FuGENE to each transfection reaction.

Large scale cell culture: Add 12 µl FuGENE to each transfection reaction.

- 4. Add the entire DNA/media/FuGENE mixture drop wise to each cell culture plate. Incubate on a shaking platform at 100 rpm for 2 minutes to evenly distribute the transfection mixture.
- 5. Return plate to humidified incubator for 24 hours.
- 6. 24 hours post-transfection, passage cells.

**Small scale cell culture:** Transfer cells in each well of a 6-well plate to a 10 cm dish.

Large scale cell culture: Transfer cells from a 10 cm dish to a 15 cm dish.

- 7. Return plate to humidified incubator for 24 hours.
- 8. Transfected cells are now ready to be processed for chromatin fixation using the enChIP Kit (Catalog No. 53125). Alternatively, if duplicate reactions were performed cell lysates can be prepared using Active Motif's Nuclear Extraction Kit (Catalog No. 40010) for Western blot analysis of transfection efficiency using the AM-Tag polyclonal antibody (Catalog No. 61677) to recognize the tagged dCas9 protein.

## **Technical Services**



If you need assistance at any time, please call Active Motif Technical Service at one of the numbers listed below.

#### **Active Motif North America**

1914 Palomar Oaks Way, Suite 150 Carlsbad, CA 92008

USA

Toll Free: 877 222 9543 Telephone: 760 431 1263 Fax: 760 431 1351

E-mail: tech service@activemotif.com

### **Active Motif Europe**

Avenue Reine Astrid, 92 B-1330 La Hulpe, Belgium

UK Free Phone: 0800 169 31 47 France Free Phone: 0800 90 99 79

Germany Free Phone: 0800 181 99 10 Telephone: +32 (0)2 653 0001 Fax: +32 (0)2 653 0050

E-mail: eurotech@activemotif.com

## **Active Motif Japan**

Azuma Bldg, 7th Floor 2-21 Ageba-Cho, Shinjuku-Ku Tokyo, 162-0824, Japan Telephone: +81 3 5225 3638 Fax: +81 3 5261 8733

E-mail: japantech@activemotif.com

## **Active Motif China**

787 Kangqiao Road Building 10, Suite 202, Pudong District

Shanghai, 201315, China

Telephone: (86)-21-20926090 Hotline: 400-018-8123

Visit Active Motif on the worldwide web at http://www.activemotif.com

## At this site:

- Read about who we are, where we are, and what we do
- Review data supporting our products and the latest updates
- Enter your name into our mailing list to receive our catalog, MotifVations newsletter and notification of our upcoming products
- Share your ideas and results with us
- View our job opportunities

Don't forget to bookmark our site for easy reference!